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## (12) United States Patent

Cooney et al.

# (54) APPARATUS FOR THE EXTRACORPOREAL TREATMENT OF BLOOD

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C12N 11/14 (2006.01)

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**A61L 33/00** (2006.01) **A61M 1/34** (2006.01)

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### (58) Field of Classification Search

None

See application file for complete search history.

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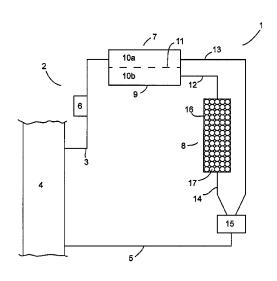
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### (57) ABSTRACT

An apparatus for the extracorporeal treatment of blood comprising an extracorporeal blood circuit (2), a pump (6) configured to provide fluid displacement within the extracorporeal blood circuit, and a reaction chamber (8) connected to the extracorporeal blood circuit and configured to receive blood or plasma from the circuit and treat the blood or plasma. The reaction chamber comprises an protease enzyme immobilized to a support, in which the protease enzyme is specific for, and capable of irreversibly cleaving, a human C5a present in the blood or plasma, wherein the abundance of the human C5a in the treated blood or plasma is less than that in the untreated blood or plasma. The apparatus finds utility in the extracorporeal treatment of blood from patients with inflammatory conditions, especially auto-immune disease and sepsis.

### 13 Claims, 2 Drawing Sheets



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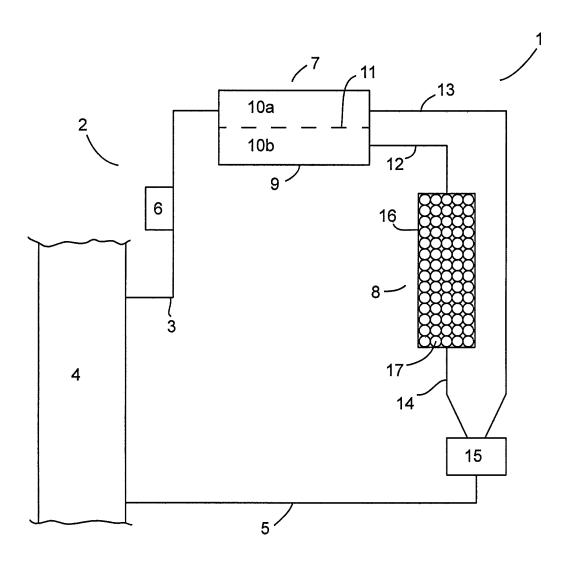
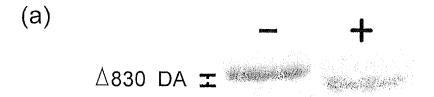


Fig. 1



(b) P4 P3 P2 P1 V P1' P2' N64s 165s S66s H67s ... K68s D69s

Fig. 2

# APPARATUS FOR THE EXTRACORPOREAL TREATMENT OF BLOOD

## CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority from European Patent Application No. 13197790.2, filed on Dec. 17, 2013, the contents of this application are hereby incorporated by reference in its entirety.

### BACKGROUND OF THE INVENTION

State-of-the-art hospital treatment for sepsis is the implementation of 'The Sepsis Six' (PMID 21398303). These are a series of interventions to stabilize the patient, including delivery of antibiotics, microbial culture, delivery of high-flow oxygen, and fluids. To date, interventions to mitigate organ damage in sepsis have failed. Treatment with Drotrecogin alfa 20 activated, a serine protease involved in switching off coagulation, was, until very recently, the major FDA-approved intervention for treatment of human sepsis. However in 2011 FDA announcing that Eli Lilly had withdrawn Xigris (Drotrecogin alfa). On the 8 Aug. 2012, AstraZeneca 25 announced that a Phase IIb study testing the efficacy of Cyto-Fab<sup>TM</sup>, an anti-TNFα, polyclonal antibody fragment, for treatment of severe sepsis and/or septic shock, did not show any significant improvement over placebo and AZ halted any further developments.

Two additional treatments have been proposed based on a blood purification strategy with some similarity to that proposed in this document. Cytosorb's IL-8 adsorption cassette is based on a porous material that adsorbs the cytokine IL-8, but the technique is non-selective, and removes other small protein components of the blood (found on the world wide web at cytosorbents.com/tech.htm). The second strategy is a specific adsorption resin removing bacterial LPS from blood circulated through a cassette (found on the world wide web at altecomedical.com/market\_product.php), and is a treatment flimited to sepsis caused by Gram negative bacteria.

There is a large body of evidence establishing the role of C5a is sepsis. The Cell Envelope Protease ScpA targets the immune proinflammatory mediator C5a and specifically cleaves the mediator rendering it active.

It is an object of the invention to overcome at least one of the above-referenced problems.

### SUMMARY OF THE INVENTION

The invention is based on a method and device for the extracorporeal treatment of inflammatory conditions in a patient, especially auto-immune diseases, sepsis or septicemia, that involves reacting blood that has been removed from a patient with a protease enzyme immobilized to a support in 55 which the enzyme is specific for a pro-inflammatory mediator present in the blood of the patient and is capable of cleaving the pro-inflammatory mediator and thereby reducing the abundance of pro-inflammatory mediator in the blood of the patient prior to the return of the treated blood to the patient. 60

In a first aspect, the invention relates to an apparatus for the extracorporeal treatment of blood comprising:

an extracorporeal blood circuit;

optionally, a pump configured to provide fluid displacement within the extracorporeal blood circuit; and

a reaction chamber connected to the extracorporeal blood circuit and configured to receive blood or a pro-inflam2

matory mediator containing blood fraction from the circuit and treat the blood or pro-inflammatory mediator containing blood fraction,

characterized in that the reaction chamber comprises a protease enzyme irreversibly immobilized to a support, in which the protease enzyme is specific for, and capable of irreversibly cleaving, a human pro-inflammatory mediator present in the blood or plasma such that the chemoattractant capability of the pro-inflammatory mediator is reduced or preferably abrogated, wherein the abundance of functional pro-inflammatory mediator in the treated blood or plasma is less than that in the untreated blood or plasma.

Compared with extracorporeal treatment devices that operate on the basis of adsorption of pro-inflammatory mediators, the apparatus of the invention has a number of advantages. Each molecule of enzyme can cleave a large number of molecules of substrate during a treatment operation; this contrasts with the adsorption process in which the ligand, once bound to its target molecule, is unavailable for binding with further target molecules. Second, the affinity antibody-based approaches of the prior art are susceptible to cross-reacting with non-target molecules, and involve significant costs in the development and generation of suitable antibodies. In contracts, enzymes that are specific to pro-inflammatory mediators are known from the literature, and can be easily produced using recombinant DNA technology.

Preferably, the pro-inflammatory mediator is selected from a group consisting of, but not limited to: C3a, C4a, C5a, IL-8, IL-6, TNF $\alpha$ , IL-1, or Mig. Thus, in one embodiment, the protease enzyme is capable of cleaving a human pro-inflammatory mediator selected from a group consisting of, but not limited to, C3a, C4a, C5a, IL-8, IL-6, TNF $\alpha$ , IL-1, and Mig.

In a preferred embodiment, the invention provides an apparatus for the extracorporeal treatment of blood comprising: an extracorporeal blood circuit;

optionally, a pump configured to provide fluid displacement within the extracorporeal blood circuit; and

a reaction chamber connected to the extracorporeal blood circuit and configured to receive blood or a human C5a-containing blood fraction from the circuit and treat the blood or human C5a-containing blood fraction,

characterized in that the reaction chamber comprises a protease enzyme irreversibly immobilized to a support, in which the protease enzyme is specific for, and capable of irreversibly cleaving, human C5a present in the blood or blood fraction such that the chemoattractant capability of the cleaved human C5a is reduced, wherein the abundance of the functional human C5a in the treated blood or blood fraction is less than that in the untreated blood or blood fraction.

As used herein, the term "functional human C5a" should be understood to mean human C5a having chemoattractant capability as determined using the chemoattractant capability assay described below. Likewise, the term "non-functional human C5a" should be understood to mean cleaved C5a protein that has reduced, or is devoid of, chemoattractant capability as determined using the chemoattractant capability assay described below.

The invention also provides an apparatus for treating human blood or a pro-inflammatory mediator-containing blood fraction, the apparatus comprising a protease enzyme irreversibly bound to a support, in which the protease enzyme is specific for, and capable of irreversibly cleaving, a pro-inflammatory mediator present in the blood or blood fraction such that the chemoattractant capability of the cleaved human pro-inflammatory mediator is reduced.

The invention also provides an apparatus for treating human blood or a C5a-containing blood fraction, the appara-

tus comprising a protease enzyme irreversibly bound to a support, in which the protease enzyme is specific for, and capable of irreversibly cleaving, human C5a present in the blood or blood fraction such that the chemoattractant capability of the cleaved human C5a is reduced.

The invention also provides a protease enzyme comprising the sequence of A-B-C-D, in which:

A is a protease enzyme that is specific for, and capable of irreversibly cleaving, a human pro-inflammatory mediator present in the blood such that the chemoattractant capability of the cleaved pro-inflammatory mediator is reduced, B is a poly-lysine, poly-cysteine or polyglutamate motif, C is a spacer (for example a short peptide of 2 to 20 amino acids), and D is a poly-histidine 15

Preferably, the protease enzyme is a recombinant bacterial C5a protease comprising a sequence of SEQUENCE ID NO: 3 or a functional variant thereof, typically having at least 70%, 80% or 90% sequence identity with SEQUENCE ID NO: 3. 20 SEQUENCE ID NO: 1 below:

The term "functional variant" as applied to SEQUENCE ID NO: 3 means a protease that is specific for, and capable of irreversibly cleaving, human C5a such that the chemoattractant capability of the cleaved human C5a is reduced, or preferably abrogated.

Examples functional variants of SEQUENCE ID NO: 3 are selected from SEQUENCE ID NO: 4 and SEQUENCE ID NO:5.

In one embodiment, the apparatus of the invention includes separating means adapted to separate the blood into a C5a-30 containing fraction and a non-05a containing fraction, wherein the reaction chamber receives the C5a-containing fraction. The separating means could be, for example, a filter configured to separate the blood or a fraction thereof into a low-molecular weight containing fraction and a second frac- 35 tion, wherein the low molecular weight containing fraction is the C5a containing fraction.

Suitably, the apparatus of the invention includes means configured to recombine the treated C5a-containing fraction (i.e. the low molecular weight fraction) with the second non- 40 05a containing fraction. The recombined fractions are then returned to the patient.

In one preferred embodiment of the invention, a C-terminal of the protease enzyme comprises a first tag and a second tag located distally of the first tag and separated from the first tag 45 by a spacer. Typically, the support comprises a coordinated transition metal ion and one or more functional groups. Suitably, the first tag comprises a motif capable of covalently reacting with the one or more functional groups, and wherein the second tag comprises a motif capable of interacting with 50 TGATAGCTTTGGGGGCAAAACCCGTCTACCTCTAGCAGATCATCCTG the coordinated transition metal ion. In this manner, the protease enzyme can be oriented with respect to the surface such that the C-terminus of the enzyme is disposed adjacent to the surface (this is achieved by the interaction between the second tag and the coordinated transition metal of the support 55 AGACGATCATCAAGCTAAAGAAATGCCTGTTCTTTCAACAAACCGTTTTG surface), thus allowing the adjacent first tag to covalently bind to the functional groups on the surface. This will prevent unspecific binding between functional groups on the surface and lysine residues in the protease enzyme.

Preferably, the first tag is selected from poly-lysine, poly- 60 glutamate, or poly-cysteine tag, and the functional groups on the surface are groups that are capable of covalently binding with these motifs.

Suitably, the second tag comprises a poly-histidine tag or another tag capable of interaction with a transition metal.

Preferably, the coordinated transition metal ion is selected from Ni<sup>2+</sup>, Co<sup>2+</sup>, Zn<sup>2+</sup>, and Cu<sup>2+</sup>.

Both tags can be appended onto the DNA sequence by PCR based methods using an oligonucleotide synthesized to contain the required sequence.

Typically, the support comprises a silica material, preferably a mesoporous silica material, preferably modified monodispersed mesoporous silicate material, and ideally a Ni<sup>2+</sup>modified mesoporous silica material. Other potential materials for the support include but are not limited to methacrylates, polyacrylamides, polypyrroles and polysaccharides.

Suitably, the support comprises a bead. Preferably, the reaction chamber comprises a column containing a multiplicity of beads.

The invention also relates to an apparatus of the invention for use in a method for the ex-vivo treatment of blood in a mammal, typically a human. Preferably, the mammal has an inflammatory condition such as sepsis.

The nucleic acid sequence encoding the bacterial C5a proprotease, ScpA from Streptococcus pyogenes, is provided in

DNA sequence (SEQUENCE ID NO: 1) GGATCCAATACTGTGACAGAAGACACTCCTGCTACCGAACAAGCCGTAGA AACCCCACAACCAACAGCGGTTTCTGAGGAAGCACCATCATCATCAAAGG AAACCAAAATCCCACAAACTCCTGGTGATGCAGAAGAAACAGTAGCAGAT GACGCTAATGATCTAGCCCCTCAAGCTCCTGCTAAAACTGCTGATACACC AGCAACCTCAAAAGCGACTATTAGGGATTTGAACGACCCTTCTCAGGTCA  $\verb|AAACCCTGCAGGAAAAAGCAGGCAAGGGAGCTGGGACTGTTGTTGCAGTG|$ ATTGATGCTGGTTTTGATAAAAATCATGAAGCGTGGCGCTTAACAGACAA AACTAAAGCACGTTACCAATCAAAAGAAGATCTTGAAAAAAGCTAAAAAAG AGCACGGTATTACCTATGGCGAGTGGGTCAATGATAAGGTTGCTTATTAC CACGATTATAGTAAAGATGGTAAAACCGCTGTCGATCAAGAGCACGGCAC ACACGTGTCAGGGATCTTGTCAGGAAATGCTCCATCTGAAACGAAAGAAC CTTACCGCCTAGAAGGTGCGATGCCTGAGGCTCAATTGCTTTTGATGCGT GTCGAAATTGTAAATGGACTAGCAGACTATGCTCGTAACTACGCTCAAGC TATCAGAGATGCTGTCAACTTGGGAGCTAAGGTGATTAATATGAGCTTTG GTAATGCTGCACTAGCTTACGCCAACCTTCCAGACGAAAACCAAAAAAGCC TTTGACTATGCCAAATCAAAAGGTGTTAGCATTGTGACCTCAGCTGGTAA  $\tt ATTATGGGGTGGTTGGGACGCCTGCAGCGGCAGACTCAACATTGACAGTT$ GCTTCTTACAGCCCAGATAAACAGCTCACTGAAACTGCTACGGTCAAAAC AGCCAAACAAGGCTTACGACTATGCTTATGCTAATCGTGGGATGAAAGAA GATGATTTTAAGGATGTCAAAGGCAAAATTGCCCTTATTGAACGTGGTGA TATTGATTTCAAAGATAAGATTGCAAACGCTAAAAAAGCTGGTGCTGTAG GGGTCTTGATCTATGACAATCAAGACAAGGGCTTCCCGATTGAATTGCCA  ${\tt AATGTTGATCAGATGCCTGCGGCCTTTATCAGTCGAAAAGACGGTCTCTT}$ ATTAAAAGACAATTCTAAAAAAACCATCACCTTCAATGCGACACCTAAGG

TATTGCCAACAGCAAGTGACACCAAACTAAGCCGCTTCTCAAGCTGGGGT

-continued

TTGACAGCTGACGGCAATATTAAGCCAGATATTGCAGCACCCGGCCAAGA TATTTTGTCATCAGTGGCTAACAACAAGTATGCCAAACTTTCTGGAACTA GTATGTCTGCGCCATTGGTAGCGGGTATCATGGGACTATTGCAAAAGCAA TATGAGACACAGTATCCTGATATGACACCATCAGAGCGTCTTGATTTAGC TAAAAAAGTATTGATGAGCTCAGCAACTGCCTTATATGATGAAGATGAAA AAGCTTATTTTCTCCTCGCCAACAAGGAGCAGCAGCAGTCGATGCTAAA AAAGCTTCAGCAGCAACGATGTATGTGACAGATAAGGACAATACCTCAAG CAAGGTTCACCTGAACAATGTTTCTGATAAATTTGAAGTAACAGTAACAG TTCACAACAATCTGATAAACCTCAAGAGTTGTATTACCAAGCAACTGTT CAAACAGATAAAGTAGATGGAAAACACTTTGCCTTGGCTCCTAAAGCATT GTATGAGACATCATGGCAAAAAATCACAATTCCAGCCAATAGCAGCAAAC CAAATGAAAAATGGCTATTTCTTAGAAGGTTTTGTTCGTTTCAAACAAGA TCCTAAAAAAAAAAAGAAGACTTATGAGCATTCCATATATTGGTTTCCGAGGTG ATTTTGGCAATCTGTCAGCCTTAGAAAAACCAATCTATGATAGCAAAGAC GGTAGCAGCTACTATCATGAAGCAAATAGTGATGCCAAAGACCAATTAGA TGGTGATGGATTACAGTTTTACGCTCTGAAAAATAACTTTACAGCACTTA CCACAGAGTCTAACCCATGGACGATTATTAAAGCTGTCAAAGAAGGGGTT GAAAACATAGAGGATATCGAATCTTCAGAGATCACAGAAACCATTTTTGC  ${\tt AGGTACTTTTGCAAAACAAGACGATGATAGCCACTACTATATCCACCGTC}$  ${\tt ACGCTAATGGCAAACCATATGCTGCGATCTCTCCAAATGGGGACGGTAAC}$ AGAGATTATGTCCAATTCCAAGGTACTTTCTTGCGTAATGCTAAAAACCT TGTGGCTGAAGTCTTGGACAAAGAAGGAAATGTTGTTTGGACAAGTGAGG TAACCGAGCAAGTTGTTAAAAACTACAACAATGACTTGGCAAGCACACTT GGTTCAACCCGTTTTGAAAAAACGCGTTGGGACGGTAAAGATAAAGACGG CAAAGTTGTTGTTAACGGAACCTACACCTATCGTGTCCGCTACACTCCGA TTAGCTCAGGTGCAAAAGAACACACACTGATTTTGATGTGATTGTAGAC AATACGACACCTGAAGTCGCAACATCGGCAACATTCTCAACAGAAGATCG TCGTTTGACACTTGCATCTAAACCAAAAACCAGCCAACCGATTTACCGTG AGCGTATTGCTTACACTTATATGGATGAGGATCTGCCAACAACAGAGTAT ATTTCTCCAAATGAAGATGGTACCTTTACTCTTCCTGAAGAGGCTGAAAC AATGGAAGGCGGTACTGTTCCATTGAAAATGTCAGACTTTACTTATGTTG TTGAAGATATGGCTGGTAACATCACTTATACACCAGTGACTAAGCTATTG GAGGGCCACTCTTAA

The amino acid sequence of the bacterial C5a pro-protease, ScpA from *Streptococcus pyogenes*, is provided in SEQUENCE ID NO: 2 below:

Protein sequence (SEQUENCE ID NO: 2)
GPLGSNTVTEDTPATEQAVETPQPTAVSEEAPSSSKETKIPQTPGDAEET
VADDANDLAPQAPAKTADTPATSKATIRDLNDPSQVKTLQEKASKGAGTV

6

-continued VAVIDAGFDKNHEAWRLTDKTKARYQSKEDLEKAKKEHGITYGEWVNDKV AYYHDYSKDGKTAVDQEHGTHVSGILSGNAPSETKEPYRLEGAMPEAQLL LMRVEIVNGLADYARNYAQAIRDAVNLGAKVINMSFGNAALAYANLPDET KKAFDYAKSKGVSIVTSAGNDSSFGGKTRLPLADHPDYGVVGTPAAADST  $\verb|LTVASYSPDKQLTETATVKTDDHQAKEMPVLSTNRFEPNKAYDYAYANRG|$ 10 MKEDDFKDVKGKIALIERGDIDFKDKIANAKKAGAVGVLIYDNQDKGFPI ELPNVDOMPAAFISRKDGLLLKDNSKKTITFNATPKVLPTASDTKLSRFS SWGLTADGNIKPDIAAPGQDILSSVANNKYAKLSGTSMSAPLVAGIMGLL OKOYETOYPDMTPSERLDLAKKVLMSSATALYDEDEKAYFSPRQQGAGAV DAKKASAATMYVTDKDNTSSKVHLNNVSDKFEVTVTVHNKSDKPQELYYQ ATVOTDKVDGKHFALAPKALYETSWOKITIPANSSKOVTVPIDASRFSKD 20 LLAQMKNGYFLEGFVRFKQDPKKEELMSIPYIGFRGDFGNLSALEKPIYD SKDGSSYYHEANSDAKDQLDGDGLQFYALKNNFTALTTESNPWTIIKAVK EGVENIEDIESSEITETIFAGTFAKODDDSHYYIHRHANGKPYAAISPNG DGNRDYVQFQGTFLRNAKNLVAEVLDKEGNVVWTSEVTEQVVKNYNNDLA STLGSTRFEKTRWDGKDKDGKVVVNGTYTYRVRYTPISSGAKEOHTDFDV IVDNTTPEVATSATFSTEDRRLTLASKPKTSQPIYRERIAYTYMDEDLPT 30 TEYLSPNEDGTFTLPEEAETMEGGTVPLKMSDFTYVVEDMAGNITYTPVT KLLEGHS

The amino acid sequence of mature bacterial C5a protease, ScpA from *Streptococcus pyogenes*, is provided in SEQUENCE ID NO: 3 below:

AEETVADDANDLAPQAPAKTADTPATSKATIRDLNDPSQVKTLQEKASKG AGTVVAVIDAGEDKNHEAWRLTDKTKARYOSKEDLEKAKKEHGITYGEWV NDKVAYYHDYSKDGKTAVDOEHGTHVSGTLSGNAPSETKEPYRLEGAMPE AQLLLMRVEIVNGLADYARNYAQAIRDAVNLGAKVINMSEGNAALAYANL PDETKKAFDYAKSKGVSIVTSAGNDSSFGGKTRLPLADHPDYGVVGTPAA ADSTLTVASYSPDKQLTETATVKTDDHQAKEMPVLSTNRFEPNKAYDYAY ANRGMKEDDEKDVKGKIALIERGDIDEKDKIANAKKAGAVGVLIYDNODK 50 GFPIELPNVDOMPAAFISRKDGLLLKDNSKKTITFNATPKVLPTASDTKL SRFSSWGLTADGNIKPDIAAPGQDILSSVANNKYAKLSGTSMSAPLVAGI  ${\tt MGLLQKQYETQYPDMTPSERLDLAKKVLMSSATALYDEDEKAYFSPRQQG}$  ${\tt AGAVDAKKASAATMYVTDKDNTSSKVHLNNVSDKFEVTVTVHNKSDKPQE}$ LYYQATVQTDKVDGKHFALAPKALYETSWQKITIPANSSKQVTVPIDASR FSKDLLAQMKNGYFLEGFVREKQDPKKEELMSIPYIGFRGDFGNLSALEK PIYDSKDGSSYYHEANSDAKDQLDGDGLQFYALKNNFTALTTESNPWTII KAVKEGVENI EDI ESSEITETI FAGTFAKQDDDSHYYIHRHANGKPYAAI SPNGDGNRDYVQFQGTFLRNAKNLVAEVLDKEGNVVWTSEVTEQVVKNYN 65 NDLASTLGSTRFEKTRWDGKDKDGKVVVNGTYTYRVRYTPISSGAKEQHT

20

8

-continued
DFDVIVDNTTPEVATSATESTEDRRLTLASKPKTSQPIYRERIAYTYMDE
DLPTTEYISPNEDGTFTLPEEAETMEGGTVPLKMSDFTYVVEDMAGNITY
TPVTKLLEGHS

The amino acid sequence of a first variant of mature bacterial C5a protease, ScpA from *Streptococcus pyogenes*, is provided in SEQUENCE ID NO: 4 below:

DANDLAPQAPAKTADTPATSKATIRDLNDPSQVKTLQEKASKGAGTVVAV IDAGFDKNHEAWRLTDKTKARYQSKEDLEKAKKEHGITYGEWVNDKVAYY  $\verb|HDYSKDGKTAVDQEHGTHVSGILSGNAPSETKEPYRLEGAMPEAQLLLMR|$ VEIVNGLADYARNYAQAIRDAVNLGAKVINMSFGNAALAYANLPDETKKA FDYAKSKGVSIVTSAGNDSSFGGKTRLPLADHPDYGVVGTPAAADSTLTV ASYSPDKQLTETATVKTDDHQAKEMPVLSTNRFEPNKAYDYAYANRGMKE DDFKDVKGKIALIERGDIDFKDKIANAKKAGAVGVLIYDNQDKGFPIELP NVDQMPAAFISRKDGLLLKDNSKKTITFNATPKVLPTASDTKLSRFSSWG LTADGNIKPDIAAPGQDILSSVANNKYAKLSGTSMSAPLVAGIMGLLQKQ YETQYPDMTPSERLDLAKKVLMSSATALYDEDEKAYFSPRQQGAGAVDAK KASAATMYVTDKDNTSSKVHLNNVSDKFEVTVTVHNKSDKPOELYYOATV QTDKVDGKHFALAPKALYETSWQKITIPANSSKQVTVPIDASRFSKDLLA QMKNGYFLEGFVRFKQDPKKEELMSIPYIGFRGDFGNLSALEKPIYDSKD GSSYYHEANSDAKDQLDGDGLQFYALKNNFTALTTESNPWTIIKAVKEGV ENIEDIESSEITETIFAGTFAKODDDSHYYIHRHANGKPYAAISPNGDGN RDYVQFQGTFLRNAKNLVAEVLDKEGNVVWTSEVTEQVVKNYNNDLASTL GSTRFEKTRWDGKDKDGKVVVNGTYTYRVRYTPISSGAKEOHTDFDVIVD NTTPEVATSATFSTEDRRLTLASKPKTSOPIYRERIAYTYMDEDLPTTEY ISPNEDGTFTLPEEAETMEGGTVPLKMSDFTYVVEDMAGNITYTPVTKLL

The amino acid sequence of a second variant of mature bacterial C5a protease, ScpA from *Streptococcus pyogenes*, 45 is provided in SEQUENCE ID NO: 5 below:

KTADTPATSKATIRDLNDPSQVKTLQEKASKGAGTVVAVIDAGFDKNHEA
WRLTDKTKARYQSKEDLEKAKKEHGITYGEWVNDKVAYYHDYSKDGKTAV
DQEHGTHVSGILSGNAPSETKEPYRLEGAMPEAQLLLMRVEIVNGLADYA
RNYAQAIRDAVNLGAKVINMSFGNAALAYANLPDETKKAFDYAKSKGVSI
VTSAGNDSSFGGKTRLPLADHPDYGVVGTPAAADSTLTVASYSPDKQLTE
TATVKTDDHQAKEMPVLSTNRFEPNKAYDYAYANRGMKEDDFKDVKGKIA
LIERGDIDFKDKIANAKKAGAVGVLIYDNQDKGFPIELPNVDQMPAAFIS
RKDGLLLKDNSKKTITFNATPKVLPTASDTKLSRFSSWGLTADGNIKPDI
AAPGQDILSSVANNKYAKLSGTSMSAPLVAGIMGLLQKQYETQYPDMTPS
ERLDLAKKVLMSSATALYDEDEKAYFSPRQQGAGAVDAKKASAATMYVTD
KDNTSSKVHLNNVSDKFEVTVTVHNKSDKPQELYYQATVQTDKVDGKHFA
LAPKALYETSWQKITIPANSSKQVTVPIDASRFSKDLLAQMKNGYFLEGF

#### -continued

VRFKQDPKKEELMSIPYIGFRGDFGNLSALEKPIYDSKDGSSYYHEANSD
AKDQLDGDGLQFYALKNNFTALTTESNPWTIIKAVKEGVENIEDIESSEI
TETIFAGTFAKQDDDSHYYIHRHANGKPYAAISPNGDGNRDYVQFQGTFL
RNAKNLVAEVLDKEGNVVWTSEVTEQVVKNYNNDLASTLGSTRFEKTRWD
GKDKDGKVVVNGTYTYRVRYTPISSGAKEQHTDFDVIVDNTTPEVATSAT
FSTEDRRLTLASKPKTSQPIYRERIAYTYMDEDLPTTEYISPNEDGTFTL
EPEAETMEGGTVPLKMSDFTYVVEDMAGNITYTPVTKLLEGHS

The proteases of SEQUENCE ID NO:s 2, 3, 4 and 5 are all capable of cleaving human C5a such that the chemoattractant capability of the cleaved protease is abrogated.

The amino acid sequence of C5a protein is provided in SEQUENCE ID NO: 6 below.

C5a protein (SEQUENCE ID NO: 6)
MLQKKIEEIAAKYKHSVVKKCCYDGACVNNDETCEQRAARISLGPRCIKA
FTECCVVASQLRANISHKDMQLGR

Other proteases that are specific to, and capable of cleaving, human C5a include ScpB from *Streptococcus agalactiae*, and functional variants thereof (Brown et al). Examples of protease enzymes capable of specifically cleaving IL-8 include ScpC from *Streptococcus pyogenes*, SpyCEP from *Streptococcus agalactiae* and functional variants thereof (Fritzer et al, Kaur et al, Zinkernagel et al, Sjolinder et al, and Hidalgo et al)

Examples of protease enzymes capable of specifically cleaving IL-6 include a published *Pseudomonas* enzyme which degrades it completely (Matheson et al). Also gingipains K and R seem to have degrading activity against several mediators, but lack specificity required.

Suitably, the apparatus further comprising means of separating whole blood into a plasma fraction and a cellular fraction, and means for recombining the cellular fraction with the treated plasma fraction. In a separation process, the plasma in the patient's blood is typically segregated from its remaining constituents. The separated plasma is mixed with an acetate buffer saturated with heparin. This lowers the plasma's degree of acidity (pH value) to 5.12, causing the LDL cholesterol, Lp(a) and fibrinogen to drop selectively out of the plasma. Together with the heparin additive, the separated 50 constituents form insoluble precipitates which can be removed from the plasma in a single filtration stage. Unused surplus heparin is held back in a separate adsorber, and bicarbonate ultrafiltration is used to restore the purified plasma to the physiologically acceptable level. The selectively treated, purified plasma is then remixed with the remaining blood constituents and supplied back to the patient. During H.E.L.P. apheresis, these four steps (plasma separation, precipitation with subsequent filtration, heparin adsorption and ultrafiltration) are performed by a single device, the PLASMAT Futura. Examples of devices capable of separating whole blood into a plasma fraction and a cellular fraction in extracorporeal blood circuits are known to the person skilled in the art, and include plasmaphoresis equipment (for example B Braun PLASMAT Futura) and hemodialysis equipment (for 65 example Gambro PHEONIX found on the world wide web at gambro.com/en/global/Products/Hemodialysis/Monitors/ Phoenix-dialysis-system/)

Typically, the reaction chamber comprises a column comprising beads in which the enzyme is immobilized to the beads. Alternatively, the reaction chamber may comprise a cartridge.

In a further aspect, the invention relates to a method for the treatment or prevention of an inflammatory condition in a human comprising the steps of reacting blood that has been removed from the patient, or a pro-inflammatory mediator containing fraction of the blood, with a protease enzyme immobilized to a support, in which the protease enzyme is specific for, and capable of irreversibly cleaving, a human pro-inflammatory mediator present in the blood or fraction such that the chemoattractant capability of the pro-inflammatory mediator is reduced or preferably abrogated, wherein the abundance of functional pro-inflammatory mediator in the treated blood or fraction is less than that in the untreated blood or fraction.

Typically, the human pro-inflammatory mediator is selected from the group consisting of, but not limited to, C3a,  $_{20}$  C4a, C5a, IL-8, IL-6, TNF $\alpha$ , IL-1, and Mig.

In a further aspect, the invention relates to a method for the treatment or prevention of an inflammatory condition in a human comprising the steps of reacting blood that has been removed from the patient, or a pro-inflammatory mediator 25 containing fraction of the blood, with a protease enzyme immobilized to a support, in which the protease enzyme is specific for, and capable of irreversibly cleaving, human C5a present in the blood or fraction such that the chemoattractant capability of the cleaved human C5a is reduced or preferably 30 abrogated, wherein the abundance of functional C5a in the treated blood or fraction is less than that in the untreated blood or plasma.

Suitably, the method includes the steps of separating the blood into a plasma fraction and a cellular fraction, treating 35 the plasma fraction, and then recombining the cellular fraction with the treated plasma fraction prior to returning the blood to the patient.

Alternatively, or in addition, the method includes the steps of separating the blood into a C5a containing fraction (for 40 example, a low molecular weight fraction) fraction and a second fraction, treating the C5a containing fraction, and then recombining the second fraction with the treated C5a containing fraction prior to returning the blood to the patient.

Typically, the method is carried out in a continuous fashion 45 using an extracorporeal blood circuit.

Suitably, the protease enzyme is a recombinant protein.

The invention also relates to support and a recombinant protease enzyme immobilized to the support, in which the recombinant protease enzyme comprises a C-terminal polyhistidine tag and a C-terminal poly-lysine tag, and in which the recombinant protease enzyme comprises a protease that is specific for, and capable of irreversibly cleaving, a human pro-inflammatory mediator present in the blood or plasma.

In this specification, the term "extracorporeal blood circuit" should be understood to mean an arrangement of conduits capable of removing blood from the body for treatment outside of the body and returning the thus treated blood to the body.

In this specification, the term "reaction chamber" should be 60 understood to mean a chamber adapted to receive blood or plasma from the extracorporeal blood circuit and allow contact between the blood or plasma and protease enzyme that is immobilized to a support within the reaction chamber.

In this specification, the term "plasma" should be under- 65 stood to mean blood from which cells have been fully or partially removed.

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In this specification, the term "pro-inflammatory mediator" should be understood to mean a host proteinaceous entity produced in the auto-immune or sepsis response which stimulates other components of the host immune system, in particular causing migration or stimulation of leukocytes of any class and progenitor forms of these cells. Specific examples of pro-inflammatory mediators specific to the human inflammatory response include C3a, C4a, C5a, IL-8, IL-6, TNFα, IL-1, and Mig.

In the specification, the term "protease enzyme that is specific for a human pro-inflammatory mediator" should be understood to mean an enzyme with the capacity to selectively, or ideally solely, break peptide bonds of pro-inflammatory mediators of human origin by hydrolysis. The protease may also be derived from the parent protease, and modified to include a functionalization group, for example one or more of a poly-histidine, poly-lysine, or poly-glutamic acid tag.

In this specification, the term "functional variant thereof" as applied to a specific protease enzyme should be understood to mean a variant of the protease enzyme that retains the ability to specifically bind and irreversibly cleave the target pro-inflammatory mediator such that the chemoattractant activity of the cleaved pro-inflammatory mediator is reduced or abrogated. Thus, for example, a functional variant of ScpA from Streptococcus pyogenes includes variant ScpA proteases that have the ability to specifically bind and irreversibly cleave the human C5a protein such that the chemoattractant capability of the cleaved protease is reduced or abrogated, and include ScpA proteases from Streptococcus pyogenes (SE-QUENCE ID NO:2, 3, 4, 5) and other Streptococcal species. The term "variant" should be understood to mean proteins or polypeptides that have at least 70% sequence homology with the reference protease, and that are altered in respect of one or more amino acid residues. Preferably such alterations involve the insertion, addition, deletion and/or substitution of 20, 10, 5 or fewer amino acids, more preferably of 4 or fewer, even more preferably of 3 or fewer, most preferably of 1 or 2 amino acids only. Insertion, addition and substitution with natural and modified amino acids is envisaged. The variant may have conservative amino acid changes, wherein the amino acid being introduced is similar structurally, chemically, or functionally to that being substituted. Typically, proteins which have been altered by substitution or deletion of catalyticallyimportant residues will be excluded from the term "variant". For any given protease enzyme, details of such catalyticallyimportant residues will be well known to those skilled in the art. Generally, the variant will have at least 70% amino acid sequence homology, preferably at least 80% sequence homology, more preferably at least 90% sequence homology, and ideally at least 95%, 96%, 97%, 98% or 99% sequence homology with the reference protease. In this context, sequence homology comprises both sequence identity and similarity, i.e. a polypeptide sequence that shares 90% amino acid homology with wild-type bacterial mature C5a peptidase is one in which any 90% of aligned residues are either identical to, or conservative substitutions of, the corresponding residues in wild-type bacterial C5a peptidase. Substitution may be conservative or non-conservative substitution, and may involve use of natural amino acids or amino acid analogues.

The term "variant" is also intended to include chemical derivatives of a protease, i.e. where one or more residues of a protease is chemically derivatized by reaction of a functional side group. Also included within the term variant are protease molecules in which naturally occurring amino acid residues are replaced with amino acid analogues.

Proteins and polypeptides (including variants and fragments thereof) of and for use in the invention may be generated wholly or partly by chemical synthesis or by expression from nucleic acid. The proteins and peptides of and for use in the present invention can be readily prepared according to well-established, standard liquid or, preferably, solid-phase peptide synthesis methods known in the art (see, for example, J. M. Stewart et al).

In this specification, the term "inflammatory condition" means a condition in which the host mounts a response to an assault. Examples of inflammatory conditions include chronic or acute inflammatory conditions including sepsis, septic shock, systemic inflammatory response syndrome, multiple organ dysfunction syndrome, hyper-reactive airway disease, allergic reaction.

In a different aspect, the invention provides a method of attaching a molecule comprising a polyaminoacid sequence to a surface, in which the C-terminal of the protease enzyme comprises a first tag and a second tag located distally of the first tag and separated from the first tag by a spacer, and in which the support comprises a coordinated transition metal ion and one or more functional groups, and in which the first tag comprises a motif capable of covalently reacting with the one or functional groups, and wherein the second tag comprises a motif capable of interacting with the coordinated transition metal ion, the method comprising the step of reacting the molecule comprising a polyaminoacid sequence with the surface.

### BRIEF DESCRIPTION OF THE DRAWINGS

The invention description below refers to the accompanying drawings, of which:

FIG. 1 is; Diagrammatic representation of blood purifying invention.

The diagram shows the components and blood flow route envisaged for the implementation of the invention. Blood is removed from the patient and fractionated into a high protein plasma fraction and a high blood cell fraction. The former is passed over the active material (immobilized enzyme) in the 40 reaction chamber and then recombined with the latter before return to the patient. Components of the invention are labeled: 1 the overall invention, 2 the extracorporeal blood purification device, 3 blood withdrawal line, 4 patient arm, 5 blood return line, 6 pumping system, 7 blood separator, 8 reaction 45 chamber, 9 cartridge housing blood separation chambers, 10a and 10b blood separation chambers, 11 biocompatible size restrictive semi-permeable membrane, 12 line delivering protein rich plasma to reaction chamber, 13 line delivering blood cell rich fraction to mixing chamber, 14 line delivering treated 50 plasma to mixing chamber, 15 mixing chamber for blood reconstitution, 16 vessel housing active component of reaction chamber, 17 reactive material comprising immobilized enzyme irreversibly coupled to solid support material.

FIG. 2 is; Activity of ScpA against the pro-inflammatory 55 mediator C5a

Panel a shows SDS-PAGE analysis of C5a untreated (-) and treated (+) with ScpA Panel b shows the scissile bond in the C5a sequence confirmed by Mass Spec analysis of C5a cleaved with ScpA.

## DETAILED DESCRIPTION OF AN ILLUSTRATIVE EMBODIMENT

Referring to the FIG. 1, there is provided an apparatus for 65 the extracorporeal treatment of blood according to the invention, and indicated generally by the reference numeral 1. The

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apparatus 1 comprises an extracorporeal blood circuit 2, having a feed line 3 for withdrawing blood from a patients arm 4 for treatment and a return line 5 for returning treated blood to the patient, and an adjustable pump 6 provided in the feed line for providing blood displacement within the blood circuit 2.

The apparatus also includes a blood separator 7 and a reaction chamber 8 in the circuit 2, the separator 7 being provided upstream of the reaction chamber 8. The separator comprises a cartridge 9 having two chambers 10a and 10b separated by a semi-permeable membrane 11 adapted to allow separation of blood proteins from blood cells. The whole blood passes from the patient to the first chamber 10a, where proteins in blood plasma pass into the second chamber 10b forming a protein rich plasma fraction in the second chamber and leaving blood cells in the first chamber 10a. A tube 12 is provided to transfer the thus-formed protein rich fraction plasma from the second chamber 10b to the reaction chamber 8 where it is treated. A further tube 13 is provided to transfer the cell rich fraction from the first chamber 10a to re-join with treated plasma distally of the reaction chamber 8 at a mixing chamber 15 where the two fractions are mixed prior to being returned to the patient via the whole blood return line 5.

The reaction chamber comprises a cylindrical vessel 16 filled with functionalized support material 17 containing the immobilized enzyme, thereby providing a large surface area for the treatment of the incoming plasma. The tube 12 feeds into a top of the cylindrical vessel 16, and the plasma filters through the cylinder before exiting the vessel through a tube 14.

Mesoporous silica (MPS) materials (including but not limited to MCM, SBA, MCF and PMO type materials) are prepared using a templated synthesis method. Ideally these particles will be monodispersed in nature. The particles will have a specific particle size in the range of 0.1-50  $\mu$ m, contain nanopores with a final internal diameter in the range 8-12 nm and have a high surface area 300-800 m<sup>2</sup> g<sup>-1</sup>.

The surface characteristics of the silica nanocarriers will be modified with a range of functional groups (e.g. —NH2, —COOH, —SH) directly during synthesis of the material, or by post synthesis grafting to facilitate covalent coupling (through the poly-Gluamate or poly-Lysine or Cysteine residues respectively) of the enzyme to the surface after orientation specific adsorption.

The Ni<sup>2+</sup>-modified MPS will be prepared by attachment of 3-iodo-trimethoxypropylsilane to the silicate surface followed by reaction with cyclam and incorporation of the metal ion. This is to generate immobilization of the protease in a controlled orientation.

In use, the extracorporeal blood circuit is connected to a patient, generally an arm of a patient, and the pump is actuated to withdraw blood from the patient and pump it through the circuit. The whole blood from the patient enters the separator 7 and is separated under pressure into the two fractions. The plasma fraction is pumped from the second chamber 10bto the reaction chamber 8 where the blood percolates through the functionalized cassette bed 17. In the reaction chamber, mediator in the plasma binds to the protease enzyme that is immobilized to the support material, and is cleaved into an inactive form that is released back into the plasma leaving the immobilized enzyme free for another reaction. As a result of the plasma passing through the reaction chamber, the concentration of functional mediator in the plasma is significantly reduced. The thus treated plasma is then pumped to the mixing chamber 15 where it rejoins with the cell rich fraction to

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form whole blood that is significantly depleted of active mediator protein. The whole blood is returned to the patient via the return line 5.

It will be appreciated that the use of a separator to filter the blood prior to treatment is optional, and that the treatment of 5 whole blood in the reaction chamber forms part of the inven-

### **EXPERIMENTAL**

### Materials and Methods

C5a Peptidase Activity Assays

Recombinant C5a was produced as an N-term His-tagged fusion (HT-05a) in accordance with the method of Toth et al., 15 and chemoattractant activity was verified in an under-agarose migration assay (data not shown). The C5a-ase activity of ScpA was demonstrated in reactions consisting of 42 nM ScpA with 37 μM HT-05a, in 50 mM Tris/HCl (pH 7.5), 100 mM NaCl, and 5 mM CaCl<sub>2</sub> for 30 min at 20° C. The observed 20 Sjölinder H, Lövkvist L, Plant L, Eriksson J, Aro H, Jones A, C5a-ase activity was independent of the presence of Complete Mini EDTAfree inhibitor cocktail (Roche). Matrix-assisted laser desorption ionization time-of-flight mass spectrometry analysis of cleaved HT-C5a was performed. Results

The activity assay showed that the ScpA cleaved C5a at a single site (FIG. 2a). MS analysis indicated a loss of 830 Da, consistent with the removal of seven residues from the C terminal (FIG. 2b) which removes chemoattractant capabili-

The invention is not limited to the embodiments hereinbefore described which may be varied in construction and detail without departing from the spirit of the invention.

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Pro	Ser	Glu 515	Arg	Leu	Asp	Leu	Ala 520	Lys	ГÀа	Val	Leu	Met 525	Ser	Ser	Ala
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Pro	Ile	Asp	Ala	Ser 645	Arg	Phe	Ser	Lys	Asp 650	Leu	Leu	Ala	Gln	Met 655	Lys
Asn	Gly	Tyr	Phe 660	Leu	Glu	Gly	Phe	Val 665	Arg	Phe	Lys	Gln	Asp 670	Pro	Lys
Lys	Glu	Glu 675	Leu	Met	Ser	Ile	Pro 680	Tyr	Ile	Gly	Phe	Arg 685	Gly	Asp	Phe
Gly	Asn 690	Leu	Ser	Ala	Leu	Glu 695	Lys	Pro	Ile	Tyr	Asp 700	Ser	Lys	Asp	Gly

Ser 705	Ser	Tyr	Tyr	His	Glu 710	Ala	Asn	Ser	Asp	Ala 715	Lys	Asp	Gln	Leu	Asp 720
Gly	Asp	Gly	Leu	Gln 725	Phe	Tyr	Ala	Leu	Lys 730	Asn	Asn	Phe	Thr	Ala 735	Leu
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Trp	Thr	Ser 835	Glu	Val	Thr	Glu	Gln 840	Val	Val	Lys	Asn	Tyr 845	Asn	Asn	Asp
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Arg	Val	Arg	Tyr	Thr 885	Pro	Ile	Ser	Ser	Gly 890	Ala	Lys	Glu	Gln	His 895	Thr
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Ala	Thr	Phe 915	Ser	Thr	Glu	Asp	Arg 920	Arg	Leu	Thr	Leu	Ala 925	Ser	Lys	Pro
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Pro	Leu	Lys	Met 980	Ser	Asp	Phe	Thr	Tyr 985	Val	Val	Glu	Asp	Met 990	Ala	Gly
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Ala 1	GIU	GIU	Tnr	Val 5	Ala	Asp	Asp	АІА	Asn 10	Asp	ьeu	Ala	Pro	Gln 15	Ala
Pro	Ala	Lys	Thr 20	Ala	Asp	Thr	Pro	Ala 25	Thr	Ser	ГÀа	Ala	Thr 30	Ile	Arg
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Asp Leu Asn Asp Pro Ser Gln Val Lys Thr Leu Gln Glu Lys Ala Ser 35 40 45

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Val	Asn	Gly	Leu	Ala 165	Asp	Tyr	Ala	Arg	Asn 170	Tyr	Ala	Gln	Ala	Ile 175	Arg
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Asp	Tyr 210	Ala	Lys	Ser	Lys	Gly 215	Val	Ser	Ile	Val	Thr 220	Ser	Ala	Gly	Asn
Asp 225	Ser	Ser	Phe	Gly	Gly 230	Lys	Thr	Arg	Leu	Pro 235	Leu	Ala	Asp	His	Pro 240
Asp	Tyr	Gly	Val	Val 245	Gly	Thr	Pro	Ala	Ala 250	Ala	Asp	Ser	Thr	Leu 255	Thr
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Thr	Met	Tyr 515	Val	Thr	Asp	Lys	Asp 520	Asn	Thr	Ser	Ser	Lys 525	Val	His	Leu
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Arg	Asn 770	Ala	Lys	Asn	Leu	Val 775	Ala	Glu	Val	Leu	Asp 780	Lys	Glu	Gly	Asn
Val 785	Val	Trp	Thr	Ser	Glu 790	Val	Thr	Glu	Gln	Val 795	Val	Lys	Asn	Tyr	Asn 800
Asn	Asp	Leu	Ala	Ser 805	Thr	Leu	Gly	Ser	Thr 810	Arg	Phe	Glu	Lys	Thr 815	Arg
Trp	Asp	Gly	Lys 820	Asp	ГÀа	Asp	Gly	Lys 825	Val	Val	Val	Asn	Gly 830	Thr	Tyr
Thr	Tyr	Arg 835	Val	Arg	Tyr	Thr	Pro 840	Ile	Ser	Ser	Gly	Ala 845	Lys	Glu	Gln
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Thr 865	Ser	Ala	Thr	Phe	Ser 870	Thr	Glu	Asp	Arg	Arg 875	Leu	Thr	Leu	Ala	Ser 880
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ГÀа	Asp	Lys	Ile	Ala 325	Asn	Ala	ГÀз	Lys	Ala 330	Gly	Ala	Val	Gly	Val 335	Leu
Ile	Tyr	Asp	Asn 340	Gln	Asp	Lys	Gly	Phe 345	Pro	Ile	Glu	Leu	Pro 350	Asn	Val
Asp	Gln	Met 355	Pro	Ala	Ala	Phe	Ile 360	Ser	Arg	Lys	Asp	Gly 365	Leu	Leu	Leu
ГÀа	Asp 370	Asn	Ser	Lys	Lys	Thr 375	Ile	Thr	Phe	Asn	Ala 380	Thr	Pro	Lys	Val
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Phe	Tyr	Ala 675	Leu	Lys	Asn	Asn	Phe 680	Thr	Ala	Leu	Thr	Thr 685	Glu	Ser	Asn
Pro	Trp 690	Thr	Ile	Ile	Lys	Ala 695	Val	Lys	Glu	Gly	Val 700	Glu	Asn	Ile	Glu

Asp Ile Glu Ser Ser Glu Ile Thr Glu Thr Ile Phe Ala Gly Thr Phe 710 Ala Lys Gln Asp Asp Asp Ser His Tyr Tyr Ile His Arg His Ala Asn  $725 \hspace{1.5cm} 730 \hspace{1.5cm} 735$ Gly Lys Pro Tyr Ala Ala Ile Ser Pro Asn Gly Asp Gly Asn Arg Asp Tyr Val Gln Phe Gln Gly Thr Phe Leu Arg Asn Ala Lys Asn Leu Val Ala Glu Val Leu Asp Lys Glu Gly Asn Val Val Trp Thr Ser Glu Val Thr Glu Gln Val Val Lys Asn Tyr Asn Asn Asp Leu Ala Ser Thr Leu Gly Ser Thr Arg Phe Glu Lys Thr Arg Trp Asp Gly Lys Asp Lys Asp 815  $\,$  815 Gly Lys Val Val Val Asn Gly Thr Tyr Thr Tyr Arg Val Arg Tyr Thr 825 Pro Ile Ser Ser Gly Ala Lys Glu Gln His Thr Asp Phe Asp Val Ile 840 Val Asp Asn Thr Thr Pro Glu Val Ala Thr Ser Ala Thr Phe Ser Thr 855 Glu Asp Arg Arg Leu Thr Leu Ala Ser Lys Pro Lys Thr Ser Gln Pro Ile Tyr Arg Glu Arg Ile Ala Tyr Thr Tyr Met Asp Glu Asp Leu Pro 890 Thr Thr Glu Tyr Ile Ser Pro Asn Glu Asp Gly Thr Phe Thr Leu Pro 900 905 Glu Glu Ala Glu Thr Met Glu Gly Gly Thr Val Pro Leu Lys Met Ser Asp Phe Thr Tyr Val Val Glu Asp Met Ala Gly Asn Ile Thr Tyr Thr 935 Pro Val Thr Lys Leu Leu Glu Gly His Ser 950 <210> SEQ ID NO 5 <211> LENGTH: 943 <212> TYPE: PRT <213 > ORGANISM: Streptococcus pyogenes <220> FEATURE: <221> NAME/KEY: MISC\_FEATURE <222> LOCATION: (1) .. (943) <223> OTHER INFORMATION: ScpA protease variant <400> SEQUENCE: 5 Lys Thr Ala Asp Thr Pro Ala Thr Ser Lys Ala Thr Ile Arg Asp Leu 1  $\phantom{\bigg|}$  5  $\phantom{\bigg|}$  10  $\phantom{\bigg|}$  15 Asn Asp Pro Ser Gln Val Lys Thr Leu Gln Glu Lys Ala Ser Lys Gly 25 Ala Gly Thr Val Val Ala Val Ile Asp Ala Gly Phe Asp Lys Asn His Glu Ala Trp Arg Leu Thr Asp Lys Thr Lys Ala Arg Tyr Gln Ser Lys Glu Asp Leu Glu Lys Ala Lys Lys Glu His Gly Ile Thr Tyr Gly Glu Trp Val Asn Asp Lys Val Ala Tyr Tyr His Asp Tyr Ser Lys Asp Gly

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Gly 145	Leu	Ala	Asp	Tyr	Ala 150	Arg	Asn	Tyr	Ala	Gln 155	Ala	Ile	Arg	Asp	Ala 160
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Val	Ser	Asp	Lys	Phe	Glu	Val	Thr	Val	Thr	Val	His	Asn	Lys	Ser	Asp

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Thr Cys Glu Gln Arg Ala Ala Arg Ile Ser Leu Gly Pro Arg Cys Ile 35 \phantom{\bigg|}40\phantom{\bigg|}45\phantom{\bigg|}
Lys Ala Phe Thr Glu Cys Cys Val Val Ala Ser Gln Leu Arg Ala Asn
Ile Ser His Lys Asp Met Gln Leu Gly Arg
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What is claimed is:

- 1. An apparatus for the extracorporeal treatment of blood comprising:
  - an extracorporeal blood circuit (2);
  - a pump (6) configured to provide fluid displacement within the extracorporeal blood circuit; and
  - a reaction chamber (8) connected to the extracorporeal blood circuit and configured to receive blood or a human the blood or human C5a-containing blood fraction,
  - characterized in that the reaction chamber comprises a protease enzyme irreversibly immobilized to a support, in which the protease enzyme is specific for, and capable of irreversibly cleaving, human C5a present in the  $\hat{b}lood^{-40}$ or blood fraction such that the chemoattractant capability of the cleaved human C5a is reduced, wherein the abundance of the functional human C5a in the treated blood or blood fraction is less than that in the untreated blood or blood fraction, and the protease enzyme is a recombinant bacterial C5a protease selected from the group consisting of SEQ ID NO: 4 and SEQ ID NO: 5.
- 2. An apparatus as claimed in claim 1 further including separating means (7) adapted to separate the blood into a C5a-containing fraction and a non-C5a containing fraction, 50 wherein the reaction chamber receives the C5a-containing fraction.
- 3. An apparatus as claimed in claim 2 further including means (15) configured to recombine the treated C5a-containing fraction with the non-C5a containing fraction.
- 4. An apparatus as claimed in claim 1, wherein a C-terminal of the protease enzyme comprises a first tag and a second tag located distally of the first tag and separated from the first tag by a spacer, the support comprises a coordinated transition metal ion and one or more functional groups for covalent 60 coupling of the protease enzyme to the support, the first tag comprises a motif covalently coupled to the one or more functional groups, and the second tag comprises a motif capable of interacting with the coordinated transition metal

5. An apparatus as claimed in claim 4, wherein the support comprises a Ni<sup>2+</sup>-modified mesoporous silica material.

- 6. An apparatus as claimed in claim 4, wherein the first tag 30 is selected from poly-lysine, poly-glutamate, and poly-cysteine, and the second tag comprises poly-histidine.
  - 7. An apparatus as claimed in claim 6, wherein the support comprises a Ni<sup>2+</sup>-modified mesoporous silica material.
- 8. An apparatus as claimed in claim 1, wherein the support C5a-containing blood fraction from the circuit and treat 35 comprises a multiplicity of beads (17) and the protease enzyme is irreversibly immobilized to a surface of the beads.
  - 9. An apparatus as claimed in claim 1 for use in a method for the ex-vivo treatment of blood in a human with sepsis.
  - 10. An apparatus for treating human blood or a C5a-containing blood fraction, the apparatus comprising a protease enzyme irreversibly bound to a support, in which the protease enzyme is specific for, and capable of irreversibly cleaving, human C5a present in the blood or blood fraction such that the chemoattractant capability of the cleaved human C5a is reduced, wherein the protease enzyme is a recombinant bacterial C5a protease selected from the group consisting of SEQ ID NO: 4 and SEQ ID NO: 5.
  - 11. An apparatus as claimed in claim 10, wherein a C-terminal of the protease enzyme comprises a first tag and a second tag located distally of the first tag and separated from the first tag by a spacer, the support comprises a coordinated metal ion and one or more functional groups for covalent coupling of the protease enzyme to the support, the first tag comprises a motif covalently coupled to the one or more functional groups, and the second tag comprises a motif capable of interacting with the coordinated metal ion.
  - 12. An apparatus as claimed in claim 11, wherein the first tag is selected from poly-lysine, poly-glutamate, and polycysteine, and the second tag comprises poly-histidine.
  - 13. A protease enzyme comprising the sequence of A-B-C-D, in which A is selected from the group consisting of SEQ ID NO: 4 and SEQ ID NO: 5, B is a poly-lysine, polycysteine, or poly-glutamate motif, C is a spacer, and D is a poly-histidine motif.